

## The effect of diflubenzuron (Dimilin® 25 WP) on some non-target aquatic insect and crustacean species

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**Abstract:** The study is aimed at evaluating, under laboratory conditions, the side effects of a commercial formulation of diflubenzuron (Dimilin® 25 WP), which is an insecticide considered a chitin synthesis inhibitor, on some non-targeted aquatic species. The effect of two lethal doses of Dimilin LC<sub>10</sub> (3.9 ng/L) and LC<sub>50</sub> (16 ng/L) previously determined on fourth instar larvae of *Culex pipiens* L. 1758 were tested on some non-target aquatic species, abundant in the Lake of Birds (Northeast Algeria), a site classified under Ramsar Convention. The tested species belonged to two classes of invertebrates (Insects and Crustaceans). The insects included four species of Heteroptera: *Corixa punctata* Illiger, 1807, *Notonecta glauca* Latreille, 1802, *Anisops sardea* Latreille, 1802 and *Plea minutissima* Leach, 1817, and one species of Coleoptera *Berosus signaticollis* Charpentier, 1825. For benthic crustaceans *Daphnia magna* Straus, 1820 (Cladocera) was retained. The results showed significant mortality recorded in *C. punctata* and medium mortality for *B. signaticollis*, *A. sardea*, *N. glauca*, *P. minutissima* and *D. magna*. Three-way ANOVA indicated highly significant effects of species, dose, and time. According to the sensitivity to Dimilin, the pairwise comparison of Tukey's test indicates that the most sensitive species was *C. punctata* followed by *B. signaticollis* and the least sensitive was *N. glauca*, followed by *D. magna*; then *A. sardea* and finally *P. minutissima*.

**Keywords:** Cladocera, Coleoptera, *Daphnia magna*, Heteroptera, toxicity

### Introduction

Insects are an integral part of the aquatic macroinvertebrate community. They play an important role in the cycling of nutrients within an ecosystem by transforming plant materials into animal tissue; moreover, they act as energy sources for other trophic levels (Albertoni & Palma-Silva 2010). Some groups among insects, such as mayflies, plecopterans and whipworms, serve as bioindicators of environmental impacts (Al-Shami *et al.* 2013, 2014, Wandscheer *et al.* 2017). Crustacean species have been known as good indicators of water quality in the global context of the eutrophication of aquatic habitats due to rapid urbanization and industrialization on the one hand, and the use of agrochemicals in agro-industrial activities on the other (Soro *et*

*al.* 2020), because they have abundant populations and are distributed in various microhabitats. The group of aquatic invertebrates includes useful individuals, as well as individuals harmful to humans and animals (disease vectors); others are phytophagous (Callisto & Gonçalves 2005).

The control methods employed against these vectors are mostly chemical. The insecticides used belong to synthetic organophosphates, pyrethroids and carbamates (Becker *et al.* 2010, Hamaidia & Soltani 2014). Insecticides remain the main means of pest control despite their negative consequences for the environment. They cause, among other things, toxicity in the food chain and pollution of the surface and groundwater (Hénaut 2011). Some pesticides are applied directly in aquatic systems to

reduce the numbers of mosquito larvae (larvicides), and thereby reduce the transmission of pathogens by mosquitoes to humans and animals (Lawler *et al.* 2017). The intensive use of insecticides becomes environmentally hostile and ecologically unsafe, since the main side effect of the application is expressed by the extinction of natural enemies of mosquitoes such as Odonates, beetles and fishes in water pools.

Environmental imperatives have pushed research toward the use of natural pesticides (Tomé *et al.* 2013, Cepeda *et al.* 2014). These new products are insect growth disruptors. They include Chitin Synthesis Inhibitors (CSI), which interfere with cuticle formation (Soltani 1991, Soltani *et al.* 1993, Chebira *et al.* 2006, Berghiche *et al.* 2007, Sun *et al.* 2015).

In Algeria, diflubenzuron is widely used against pest insects in forestry. Moreover, diflubenzuron was previously found in *Penaeus kerathurus* shrimp to disturb the fine structure of the different cuticle layers (Morsli & Soltani 2003). Bioassays conducted under laboratory conditions have shown that CSI like diflubenzuron and triflumuron were found potent for mosquito control (Soltani *et al.* 1999, Soltani & Rehim 2001). Many biochemical effects of diflubenzuron have also been reported on the metabolism of carbohydrates (Soltani 1990) and lipids (Khebbab *et al.* 1997). Dimilin was reported to affect growth and glutathion activity in mosquitofish *Gambusia affinis* (Drardja-Beldi & Soltani 2003). More recently Novaluron, a CSI, was reported to affect moulting hormone, cuticle secretion and chitin contents in the shrimp *Palaemon adspersus* (Berghiche *et al.* 2018), while in the shrimp *Palaemon adspersus* the same product altered the biochemical composition of cuticle and induced oxidative stress (Lechekhab & Soltani 2018).

In this study, we analyzed the effect of diflubenzuron against a few non-target aquatic species, all considered good bioindicators. Two lethal doses of Dimilin LC<sub>10</sub>

(3.9 ng/L) and LC<sub>50</sub> (16 ng/L) on *C. punctata*; *N. glauca*; *A. sardea*; *P. minutissima*; *B. signicollis* and *D. magna*, were tested.

## Material and methods

### Sampling area

Lake of Birds (36° 47'N 08° 7'E) has a more or less oval surface with a characteristic pond tail stretching to the northwest with shallow sloping shores. It covers a total area of 70 ha with a deposit of 20 cm of organic matter (Houhamdi & Samraoui 2002) (Fig. 1). The study site was chosen due to its ecological significance as a continental freshwater aquatic site, favorable for the development of Culicidae and a wide range of invertebrate animals, and also because it is geographically located in an ecosystem protected by Ramsar Convention and notable for its rich animal and floral biodiversity.

### Biological models

The insect species tested are all considered good bioindicators. The aquatic Heteroptera is a heterogeneous taxonomic group of Hemiptera. They are a suborder of hemipterans characterized by the piercing – sucking rostrum type and partially sclerified front wings. The aquatic lifestyle provides them with a number of adaptations such as water repellent pads, natatory setae, respiratory siphons and aerial plastrons (Poisson 1957, Loncle 2020). The aquatic Coleoptera are the only holometabolous insects present in both imaginal and larval forms, characterized by the presence of a mouthpiece of the crusher type, and leathery forewings unfit for flight (Forge 1981). However, Cladocerans species are small aquatic crustaceans, with a much reduced number of segments, and thorax and abdomen fused. They move thanks to swimming movements, jerks of the well-developed antennae.

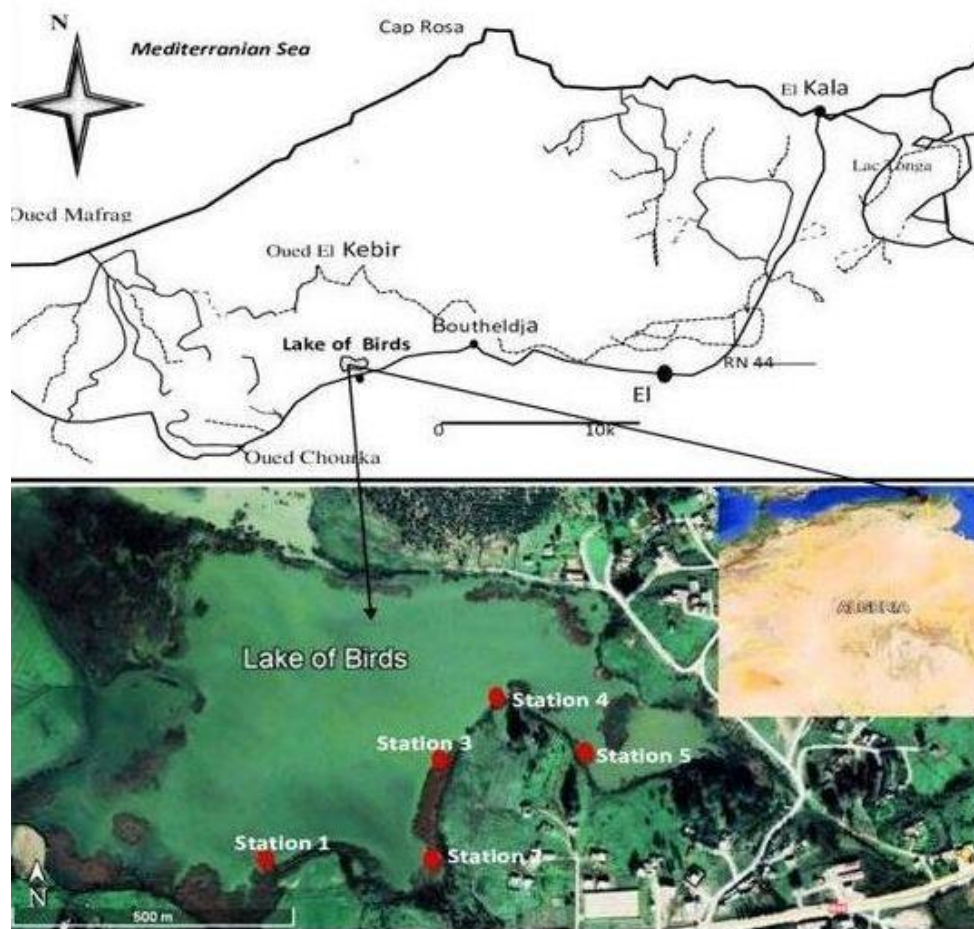


Fig. 1. Geographical location Lake of Birds and five sampling stations.

### **Insecticide and treatment**

Dimilin® (Wettable Powder, 25% active ingredient, a.i.), a commercial formulation of diflubenzuron, is an insecticide belonging to benzoylphenyl urea derivatives. The two concentrations of Dimilin, previously determined on fourth stage larvae of *Culex pipiens* L. 1758 (Rehimi 2004), were added to bioassay vessels. The first concentration test corresponds to the  $LC_{10}$  (3.9 ng/L) and the second to the  $LC_{50}$  (16 ng/L). The two concentrations were tested on the six species present in abundance at the study site (*C. punctata*; *A. sardea*; *N. glauca*; *P. minutissima*; *B. signaticollis* and *D. magna*). The test was carried out in plastic boxes containing 250 ml of lake water and food (mosquito larvae) under laboratory conditions

at a temperature of 24°C and 76% humidity. Each test consisted of three controls and three repetitions for each dose, tested on the six species; 20 individuals were exposed to these concentrations in each repetition of each species. Mortality was recorded 24 h, 48 h, and 72 h after treatment.

### **Data analysis**

All statistical analyses were performed using R, version 4.0.1 (R Core Team 2020). The results are given as the means  $\pm$  SD (standard deviation), variations regarding the percentage of mortality are plotted through the histogram. Two-way ANOVA was used to analyze the variation in mortality between the species for each exposure time. Three-way ANOVA was used to test the variance in individual deaths, according to species, dose

and time. In box plots graphs and comparison of Tukey's HSD test between times and between studied species using the package 'ggplot2' (Wickham 2016) different lowercase letters indicate a significant difference between the levels of the studied factors. All the statistical analyses were conducted at  $\alpha=0.05$  as a significance level.

## Results

The sensitivity to the insecticide was highly significant between species ( $p < 0.001$ ), depending on the time of exposure.

Dimilin at lower dose (3.9 ng/L) showed a toxic effect on the six species. According to the mortality rate, the species are classified in descending order (Fig. 2). At  $LC_{10}$ -after 24 h of

exposure, *B. signaticollis* was the most sensitive with  $2.66 \pm 0.57\%$  mortality, followed by *C. punctata* ( $2.33 \pm 0.6\%$ ), *N. glauca* ( $2.33 \pm 0.6\%$ ), *D. magna* ( $2.0 \pm 0.57\%$ ), *P. minutissima* ( $1.33 \pm 0.1 \%$ ) and *A. sardeae* ( $0.66 \pm 0.1\%$ ). After 48h of exposure, mortality was estimated at  $7.33 \pm 0.5\%$  to *C. punctata*, followed by *N. glauca* ( $4.33 \pm 0.6\%$ ), *B. signaticollis* ( $4.00 \pm 0.15\%$ ), *A. sardeae* ( $3.66 \pm 1.73\%$ ), *D. magna* ( $2.33 \pm 0.5\%$ ) then *P. minutissima* ( $2.33 \pm 0.2\%$ ).

After 72h of exposure, *C. punctata* was still the most sensitive to Dimilin with  $11.33 \pm 1.8\%$  mortality, followed by *N. glauca* ( $7.66 \pm 1.2\%$ ), *B. signaticollis* ( $6.33 \pm 0.57\%$ ), *A. sardeae* ( $4.66 \pm 0.2\%$ ), *P. minutissima* ( $4.33 \pm 0.57\%$ ) and *D. magna* ( $4.0 \pm 1\%$ ).

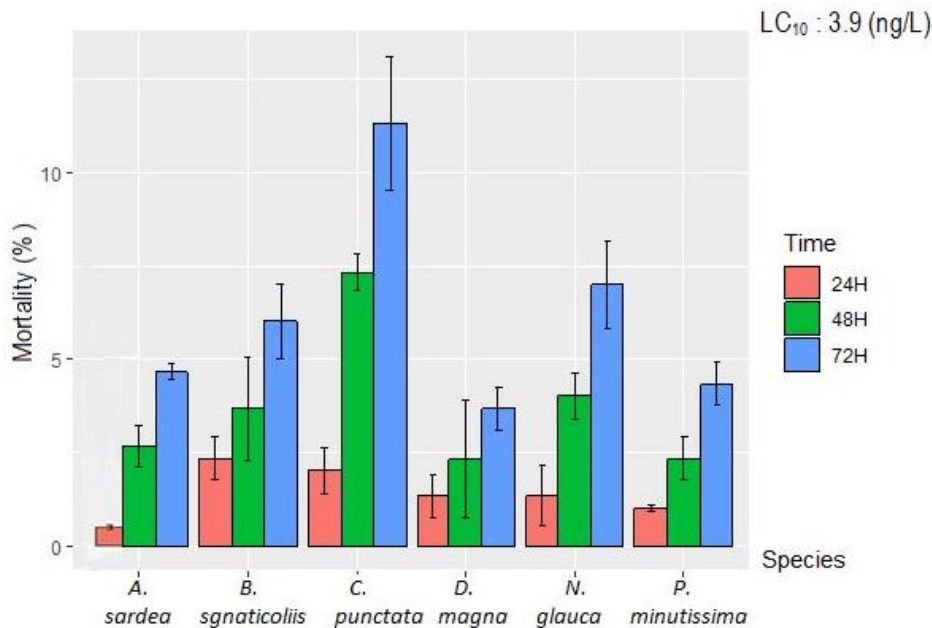


Fig. 2. Mortality rate (%) of the six species treated with Dimilin at  $LC_{10}$  during the three exposure times.

For the higher dose of Dimilin (16 ng/L) the results show a highly significant ( $p < 0.001$ ) sensitivity depending on the species and time of exposure (Fig. 3). At  $LC_{50}$ , after 24h of exposure, *C. punctata* was the most sensitive species with  $9.0 \pm 0.8\%$  mortality followed by *B. signaticollis* ( $4.0 \pm 0.2\%$ ), *D. magna* ( $3.0 \pm 1\%$ ), *N. glauca* ( $3.0 \pm 0.1\%$ ), *A. sardeae* ( $2.66 \pm$

$0.4 \%$ ) then *P. minutissima* ( $1.0 \pm 0.6\%$ ). After 48h of exposure, mortality increased significantly for *C. punctata* ( $13.66 \pm 1.2\%$ ), then *B. signaticollis* ( $7.66 \pm 0.57 \%$ ), *N. glauca* ( $5.33 \pm 1\%$ ) and *D. magna* ( $4.33 \pm 0.57 \%$ ), *P. minutissima* ( $4.0 \pm 1\%$ ) and *A. sardeae* ( $3.66 \pm 0.5 \%$ ). After 72h of exposure, *C. punctata* was the most sensitive with  $17 \pm 1.5\%$  mortality,

followed by *B. signaticollis* ( $12.33 \pm 1.0\%$ ), *D. magna* ( $7.66 \pm 1.15\%$ ), *N. glauca* ( $7.33 \pm 1.25\%$ ), *P. minutissima* ( $6.0 \pm 1.0\%$ ) and *A. sardea* ( $6.0 \pm 1.0\%$ ).

The three-way analysis of variance showed that the toxicological effect of Dimilin was significantly dependent on species ( $F_{99} = 40.71$ ;  $p = 0.000$ ), dose ( $F_{99} = 55.36$ ;  $p = 0.000$ ) and exposure time ( $F_{99} = 85.50$ ;  $p = 0.000$ ).

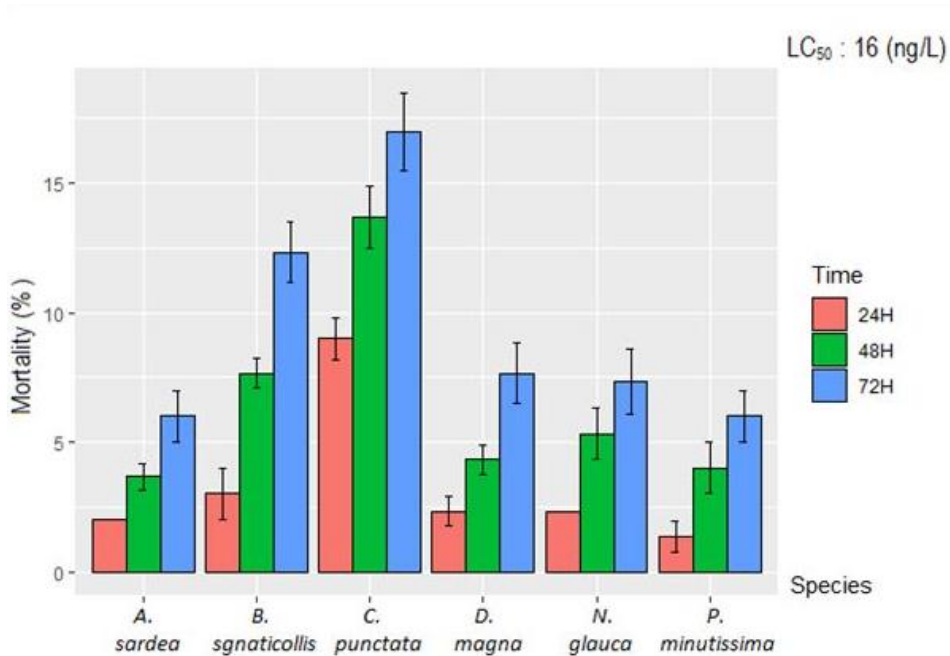


Fig. 3. Mortality rate (%) of the six species treated with Dimilin at LC<sub>50</sub> during the three exposure times.

The pairwise comparison of Tukey's test indicates that the box plots of the variation between time revealed the existence of highly

significant differences, a single group for 24 h and 72 h, and 2 homogeneous groups for 48 h (Fig. 4).

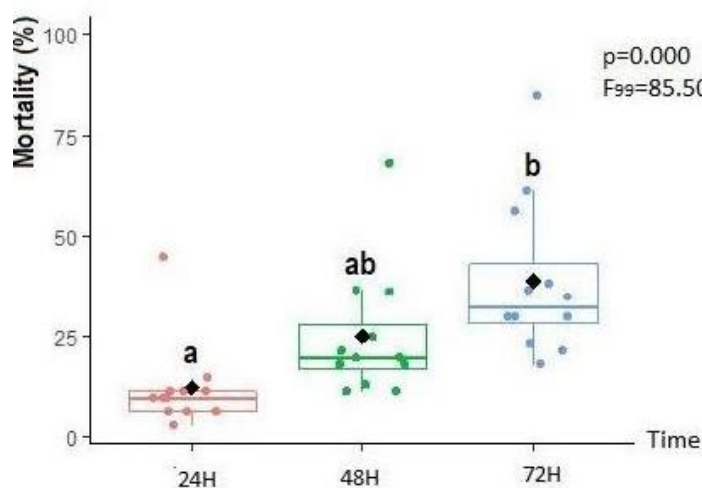


Fig. 4. Time variations regarding the percentage of mortality under the effect of Dimilin. The whisker boxes labeled with the same letter are not significantly different at  $p > 0.05$  (Tukey's test). The center box boundaries show the interquartile range (IQR) with the first quartile (lower bound) and third quartile (upper bound).

While analyzing the box plots of the species factor, we observed significant differences in the studied species, with clear heterogeneity, two groups homogeneous for

*B. signaticollis* and only one group for *A. sardea*, *C. punctata*, *N. glauca*, *P. minutissima* and *D. magna* (Fig. 5).

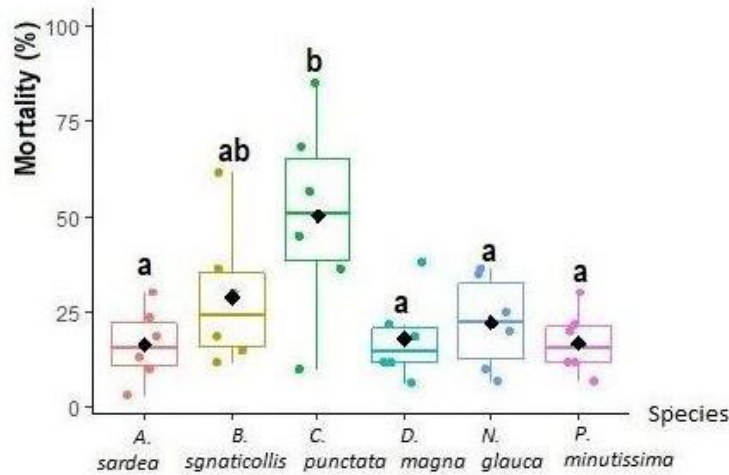


Fig. 5. Species variations regarding the percentage of mortality under the effect of Dimilin. The whisker boxes labeled with the same letter are not significantly different at  $p > 0.05$  (Tukey's test). The center box boundaries show the interquartile range (IQR) with the first quartile (lower bound) and third quartile (upper bound).

## Discussion

The data obtained from this experiment indicates that Dimilin has a negative effect on associated insect fauna. In our work, the treatment of six species of invertebrates resulted in variable sensitivity to Dimilin ( $LC_{10}$ : 3.9ng/L and  $LC_{50}$ : 16ng/L). The results show significant toxicity recorded in *C. punctata*; *B. signaticollis* and average toxicity for *N. glauca*; *D. magna*; *P. minutissima* and *A. sardea*.

The insecticides play a great role in the management of insect pests, however the concentration and indiscriminate use of treatment methods are criticized in view of their negative effect on some non-target aquatic insects., and adverse effects on soil characteristics (Khudhur & Sarmamy 2019). Benzoylphenyl urea derivatives interfere with the molting process by disrupting cuticular secretion via chitin synthesis (Morsli & Soltani 2003, Soltani *et al.* 2009, Berghiche *et al.* 2018). The detrimental effects of CSI are felt

during various critical phases of insect development. It was also reported that Novaluron, another inhibitor of chitin synthesis, caused a decrease in the level of cuticular chitin of *P. adspersus* (Berghiche *et al.* 2016).

Diflubenzuron is an insecticide that has been widely used for the selective control of insect pests (Subrero *et al.* 2018). It was first discovered as a post-ingestion larvicide, but further studies determined that this insecticide could also prevent egg hatching after direct egg contact or after the female treatment method (Singh 2015). The sensitivity of *D. magna* to diflubenzuron could be related to inhibition of chitin synthesis and chitinase activity (Kota *et al.* 2022). Subrero *et al.* (2018) determined acute toxicity following exposure to the diflubenzuron (0.15; 0.015; 0.0015 mg/L) on egg hatching rate and motility inhibition of *D. magna*. Similarly, diflubenzuron showed high toxicity to *D. magna*, indicating that the use of this

substance could lead to a high environmental risk for this species (Abe *et al.* 2014).

Dimilin in dose of 16 ng/L and 1 ng/L applied to the crustacean of type *P. adspersus* shrimp induced a significant decrease in the amounts of chitin at the treated doses after 96 h (Lechekhab & Soltani 2018). This product may have side effects on non-target arthropods such as shrimps, since a similar effect has been reported on another shrimp *Penaeus kerathurus* (Soltani *et al.* 2009). According to Macken *et al.* (2015), diflubenzuron interferes with enzymes that contribute to the synthesis of chitin in crustacean species such as benthic copepods *Tisbe battagliai*. Dimilin was also found to be toxic to Cladocera and Copepoda crustacean zooplankton (Lahr *et al.* 2000). A study by Harðardóttir *et al.* (2019) explained the mechanism of diflubenzuron (93.2 ng/L) action on *Lepeophtheirus salmonis* (Copepoda, Caligidae) and confirmed inhibition of chitin production. On the other hand, in their work Ferreira *et al.* (2020) tested the use of diflubenzuron in following doses 250 mg/L, 750 mg/L and 1 g/L for *Buenoa* sp. (Heteroptera: Notonectidae). A strong relationship between the mortality and exposure time in all concentrations was observed. The obtained lethal concentrations of LC<sub>50</sub> (2.77 x 10<sup>-3</sup> g/L) and LC<sub>90</sub> (0.86 g/L) for *Buenoa* sp. were below the recommended dose for mosquito control.

Our results show that Dimilin was found to exhibit significant toxicity against *C. punctata*, and average toxicity on the other studied species. In this context, there is a need for tools to monitor the toxicity risk of these pesticide to the environment, a negative effect on non-target insects and natural predators. The evaluation of the richness and the toxicology of the study site will allow us to carry out further studies concerning the use of mainly natural control products for the preservation of the environment and biological balance.

## Conclusions

Chemical control has become a source of enormous environmental problems, and alternative methods are searched. The obtained results showed that the two tested doses had a lethal effect on the studied species over time. Our results show that Dimilin was found to exhibit significant toxicity against *C. punctata*, and average toxicity to the other studied species. The evaluation of the richness and the toxicology of the study site will allow us to carry out further studies concerning the use of mainly natural control products for the preservation of the environment, the development of new biomonitoring strategies.

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## References

- Abe FR, Coleone AC, Machado AA, Goncalves Machado-Neto J. 2014. Ecotoxicity and environmental risk assessment of larvicides used in the control of *Aedes aegypti* to *Daphnia magna* (Crustacea, Cladocera). *Journal of Toxicology and Environmental Health*, 7: 37-45.
- Albertonie F, Palma-Silva C. 2010. Characterization and importance of two invertebrates of continental waters common to the environments of Rio Grande. *Cad Ecology Aquatic*, 5 (1): 9-27.
- Al-Shami AS, Heino J, Salmah MRC, Madrus MR. 2013. Drivers of beta diversity of macroinvertebrate communities in tropical forest streams. *Freshwater Biology*, 58: 1126-1137.

- Al-Shami SA, Rawi CSM, Ahmad AH, Madrus MR, Mutairi K. 2014. Importance of regional diversity and environmental conditions on local species richness of aquatic macroinvertebrates in tropical forested streams. *Journal of Tropical Ecology*, 30 (4): 335-346.
- Becker N, Petric D, Zgomba M, Dahl C, Boase C, Lane J, Kaiser A. 2010. Genetic control of mosquitoes. In: Becker N, Petric D, Zgomba M, Dahl C, Boase C, Lane J, Kaiser A. (Eds). *Mosquitoes and Their Control*. Springer, Berlin, Heidelberg, pp. 483-490.
- Berghiche H, Smagghe G, Van DVS, Soltani N. 2007. In vitro cultures of pupal integumental explants to bioassay insect growth regulators with ecdysteroid activity for ecdysteroid amounts and cuticle secretion. *African Journal of Agricultural Research*, 2 (5): 208-213.
- Berghiche H, Benradia H, Soltani N. 2016. Impact of insect growth disruptor, Novaluron, on biochemical composition of cuticle from the shrimp *Palaemon adspersus*. *Journal of Entomology and Zoology Studies*, 4 (2): 147- 151.
- Berghiche H, Benradia H, Soltani N. 2018. Evaluation of the potential side-effects of Novaluron on the shrimp *Palaemon adspersus*: moulting hormone profile, cuticle secretion and chitin contents. *International Journal of Environmental Monitoring and Analysis*, 6 (4): 116-124.
- Callisto M, Gonçalves JF. 2005. Aquatic invertebrates as bioindicators. Navigating the rio das velhas from minas to gerais. *Accelerating the world's research*, 1: 1-12.
- Cepeda-Palacios R, Servín R, Ramírez-Orduña JM, Ascencio F, Dorchie P, Angulo-Valadez, C.E. 2014. In vitro and in vivo effects of neem tree products on larvae of the sheep nose bot fly (*Oestrus ovis* L. Díptera: Oestridae). *Veterinary Parasitology*, 200: 225-228.
- Chebira S, Soltani N, Muylle S, Smagghe G. 2006. Uptake and distribution of three insect growth regulators-diflubenzuron, flucyclozuron and halofenozide- in pupae and adults of *Tenebrio molitor*. *Phytoparasitica*, 34 (2): 187-196.
- Drardja-Beldi H, Soltani N. 2003. Laboratory evaluation of Dimilin on growth and glutathion activity in mosquitofish, a non-target species. *Communications in Agricultural and Applied Biological Sciences*, 68 (4a): 299-305.
- Ferreira FA, Arcos AN, Maia NS, Sampaio R, Costa FM, Rodrigues IB, Tadei WP. 2020. Effects of diflubenzuron on associated insect fauna with Anopheles (Diptera: Culicidae) in laboratory, partial-field and field conditions in the Central Amazon. *Anais da Academia Brasileira de Ciências*, 17;92(1):e20180590. doi: 10.1590/0001-3765202020180590.
- Forge P. 1981. Beetles. Flora and aquatic fauna of Sahelo-Sudanian Africa, Determination key, ORSTOM, V II, 487 pp.
- Hamaidia K, Soltani N. 2014. Laboratory evaluation of a biorational insecticide, kinoprene, against *Culex pipiens* larvae: Effects on growth and development. *Annual Research & Review in Biology*, 4 (14): 2263-2273.
- Harðardóttir H, Male R, Nilsen F, Dalvin S. 2019. Effects of chitin synthesis inhibitor treatment on *Lepeophtheirus salmonis* (Copepoda, Caligidae) larvae. *PLoS One*, 14.
- Hénaut A. 2011. *Air and water pollution. Files of science and public policies*. Pierre and Marie Curie University, Paris, 64 pp.
- Houhamdi M, Samraoui B. 2002. Spatio - temporal occupation by the aquatic avifauna of Lac the Birds (Algeria). *Alauda*, 70 (2): 301-310.
- Khebbeb MEH, Delachambre J, Soltani N. 1997. Lipid metabolism during the sexual maturation of the mealworm (*Tenebrio molitor*): effect of ingested diflubenzuron. *Pesticide Biochemistry and Physiology*, 58 (3): 209-217.
- Khudhur NS, Sarmamy A. 2019. The Determination of diazinon residues in



- artificially polluted soils. *Zanco Journal Of Pure And Applied Sciences*, 31: 1-8.
- Kota K, Kazuyuki O, Kyoshiro H, Hiroyuki K, Keiko N, Takahiro Y, Nobuyoshi N, Haruna W, Hiroshi Y. 2022. Potential differences in chitin synthesis ability cause different sensitivities to diflubenzuron among three strains of *Daphnia magna*. *Aquatic Toxicology*, 243.
- Lahr J, Diallo A, Gadji B, Diouf P, Bedaux J, Badji A, van Straalen M. 2000. Ecological effects of experimental insecticide applications on invertebrates in Sahelian temporary ponds. *Environmental Toxicology and Chemistry: An International Journal*, 19(5): 1278-1289.
- Lawler S .2017. Environmental safety review of methoprene and bacterially-derived pesticides commonly used for sustained mosquito control. *Ecotoxicology and Environmental Safety*, 139: 335-343.
- Lechekhab H, Soltani N. 2018. Environmental risks of an insecticide (Dimilin® 25 wp) on the shrimp *Palaemon adspersus*: biochemical composition of cuticle and oxidative stress. *Fresenius environmental bulletin*, 27 (3): 1862-1867.
- Loncle P. 2020. Aquatic Heteroptera. Observatory of Continental Invertebrates of Brittany, "Taxonomic" sheets SHEET N°3, 13 pp.
- Macken A, Lillicrap A, Langford K. 2015. Benzoylurea pesticides used as veterinary medicines in aquaculture: Risks and developmental effects on non-target crustaceans. *Environmental Toxicology and Chemistry*, 34 (7): 1533-1542.
- Morsli M.S, Soltani N.2003. Effects of an insecticide chitin synthesis inhibitor, diflubenzuron on the shrimp cuticle *Penaeus kerathurus*. *Journal de Recherche Océanographique*, 28(12): 85-88.
- Poisson R. 1957. Aquatic Heteroptera. Fauna of France, 61: 1-263.
- R Core Team (2020) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.r-project.org>.
- Rehimi N. 2004. Study of reproduction in *Culex pipiens*. Aspects: morphological, ethological and physiological. Effects of some developmental inhibitors on some biological parameters. State doctoral thesis in Natural Sciences, mention Animal Biology. Badji Mokhtar University of Annaba, Algeria. PhD Thesis, 195pp.
- Singh S. 2015. Impact of New Chemistry on Biocontrol Agents of Major Crop Pests. *International Journal Of Agricultural Science And Veterinary Medicine*, 3: 14-33.
- Soltani N. 1990. Action of duflubenzuron and 20-hydroxyecdysone on carbohydrates and hemolymphatic proteins in nymphs of *Tenebrio molitor*. *Annales de la Société Entomologique de France*, 26 (4):575–584.
- Soltani N. 1991. A new point of attack: inhibition of chitin synthesis in insects. *Public Health* (Bayer AG Leverkusen, Germany), 8: 6-11.
- Soltani N, Chebira S, Delbecque JP, Delachambre J. 1993. Biological activity of flucycloxuron, a novel benzoylphenylurea derivative, on *Tenebrio molitor*: comparison with diflubenzuron and triflumuron. *Experientia*, 49 (12): 1088-1091.
- Soltani N, Lechekhab H, Smagghe G. 2009. Impact of the insect growth regulator diflubenzuron on biochemical composition of cuticle of the shrimp *Penaeus kerathurus*. *Communications in Agricultural and Applied Biological Sciences*, 74 (1): 137-141.
- Soltani N, Rehimi N. 2001. Laboratory evaluation of Andalin, a new insecticide interfering with cuticle deposition, against *Culex pipiens*. *Algerian Journal of Medicine*, 1 (XI): 28–33.
- Soltani N, Rehimi N, Drardja H, Bendali F. 1999. Activity of triflumuron against *Culex pipiens* and impacts on two non-target larvivorous species. *Annals of the Entomological Society of France*, 35: 59-64.

- Soro TA, Etile R N, Goore G Bi, Aboua B. 2020. Preliminary study of the zooplankton population in the Upper Bandama basin (Ivory Coast). *Agronomie Africaine*, 31(3): 305-319.
- Subrero E, Sforzini S, Viarengo A, Cucco M. 2018. Exposure to anti-mosquito insecticides utilized in rice fields affects survival of two non-target species, *IschnuraeI egans* and *Daphnia magna*. *Paddy and Water Environment*, 17: 1-11.
- Sun R, Liu C, Zhang H, Wang Q. 2015. Benzoylurea Chitin Synthesis Inhibitors. *Journal of agricultural and food chemistry*, 63 (31): 6847-6865.
- Tomé HVV, Martins J C, Corrêa A S, Galdino TVS, Picanço MC and Guedes RNC. 2013. Azadirachtin avoidance by larvae and adult females of the tomato leafminer *Tuta absoluta*. *Crop Protection*, 46: 63-69.
- Wandscheer ACD, Marchesan E, Santos S, Zanella R, Silva MF, Londero GP, Donato G. 2017. Richness and density of aquatic benthic macroinvertebrates after exposure to fungicides and insecticides in rice paddy fields. *Anais da Academia Brasileira de Ciências*, 89: 355- 36.
- Wickham H. 2016. *ggplot2: elegant graphics for data analysis*. Springer Nature, Switzerland, 260 pp.

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